



NordVal International Certificate

Issued for:	HyServe Compact Dry X-SA Method for the Enumeration of <i>Staphylococcus aureus</i> in Foods
NordVal No:	042
First approval date:	1 June 2011
Renewal date:	1 June 2017
Valid until:	1 June 2019

HyServe Compact Dry X-SA

Manufactured by:
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Supplied by:
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Hechenrainerstr 24,
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fulfils the requirements of the NordVal validation protocol. The reference method was ISO 6888-1:1999: Microbiology of food and animal feeding stuffs - Horizontal method for the enumeration of coagulase-positive staphylococci (*Staphylococcus aureus* and other species) – Part 1: Technique using Baird-Parker agar medium.

NordVal International has reviewed the method and the validation studies conducted by Campden BRI, UK. The studies have been conducted according to ISO 16140-2:2016. The results document no statistical difference in the performances between Compact Dry X-SA and the ISO 6888-1: 1999.

Date: 26 May 2017

Yours sincerely



Hilde Skår Norli
Chair of NordVal International



Nina Skall Nielsen
NMKL Secretary General



PRINCIPLE OF THE METHOD

Compact Dry X-SA method contains a ready-to-use dry chromogenic medium, and selective agents for the detection and enumeration of *Staphylococcus aureus*, which form blue colonies. An aliquot of 1 ml of an appropriate dilution is plated onto Compact Dry X-SA plate. The incubation conditions tested in the study were $37 \pm 1^\circ\text{C}$ for $24 \pm 2\text{h}$.

FIELD OF APPLICATION

The method has been tested on the detection and enumeration of *Staphylococcus aureus* in foods.

HISTORY

Original studies for this certificate were conducted according NordVal protocol from 2010. Selectivity results are from this study. A renewal study was carried out in 2017 according to ISO 16140-2:2016 to test for relative trueness and accuracy profile. The design of the Inter-laboratory study (ILS) is the same for the 2003 and 2016 versions of ISO16140, and therefore the existing ILS data were reanalysed using the new statistical approach outlined in ISO16140-2:2016.

RESULTS OF THE COMPARISON STUDIES

Selectivity; inclusivity and exclusivity

Inclusivity: All the 32 strains of *Staphylococcus aureus* yielded typical (blue) colonies on Compact Dry X-SA. They also appeared as typical colonies (black colonies surrounded by a zone of precipitation and halo) on Baird-Parker agar (BPA) medium.

Exclusivity: 23 strains of non-target organisms were tested. None of the strains appeared typical (blue colonies) on the Compact Dry X-SA. Four of the strains grew on BPA and yielded presumptive positive colonies. These included the coagulase positive *Staphylococcus hyicus* and *Enterococcus faecalis* and *Staphylococcus caprae* and *Staphylococcus hyicus intermedius*.

Precision and compliance between Compact Dry X-SA and the reference method

During this study it was necessary to artificially contaminate the majority of samples. The foods tested were cooked sliced ham, frozen prawns, raw cow's milk, fresh cream pastry products and chilled fresh pasta. At least five levels of contamination were used for each food matrix. Each sample was tested by both the reference method and the alternative method.

Relative trueness

The relative trueness is illustrated by the use of a Bland-Altman plot, i.e. the difference (bias) between paired samples analysed with the reference method and the alternative method respectively, plotted against the mean values obtained by the reference method. In the plot, Upper and Lower limits are included as the bias ± 2 times the standard deviation of the bias. The Bland-Altman Plot in Figure 1, illustrates the difference obtained in the enumeration of total *Staphylococcus aureus* in foods by the alternative and the reference method, respectively.

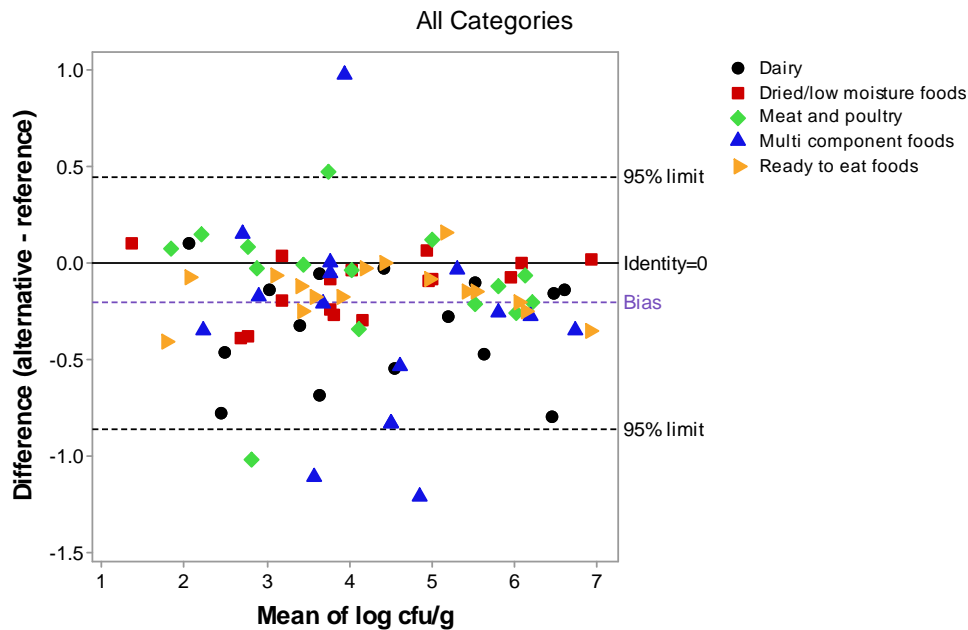


Figure 1: Bland-Altman plot for all categories

Conclusion of the comparison study:

The results of the Bland-Altman Plot provides a visual observation on the amount of bias and extreme results. It is expected that not more than one in 20 data values will lie outside the 95% Confidence Limits (CLs). The Figure 1, shows that there is a bias, as more results are obtained below the Identity =0.

For 'All Categories' there are five in 77 values which lie outside the CLs. This is a little more than the expectation of less than one in 20. For this circumstance or more extreme ($\geq 5/77$) a binomial distribution with mean = 1/20 gives a probability of 0.341, substantially greater than the commonly accepted critical significance level of 0.05. There were no identifiable trends in the five data outside the CLs and they covered 4 different food categories, 2 different inoculated strains and naturally contaminated samples.

Accuracy profiles:

The accuracy profile study is a comparative study between the results obtained by the reference and the results of the alternative method. This study is conducted using artificially contaminated samples. One type per category is tested for this.

Food matrices

For each of 5 food categories, one type of food was tested using 6 samples per type. Of the 6 samples, there were 2 at a low level, 2 at a medium level and 2 at a high level of contamination. For each of the 6 samples per category, 5 replicate test portions were tested. Non-inoculated samples (5) of each product type were also tested. Each sample was bulk inoculated and separate replicate test portions examined.



The observed profiles are within the 0.5 log AL or the recalculated AL limit calculated according to ISO16140-2:2015 section 6.1.3.3.

All the accuracy profiles, shown in figure 2, fulfil the performance criteria and the alternative method is accepted as being equivalent to the reference method.

Figure 2a: Dairy products

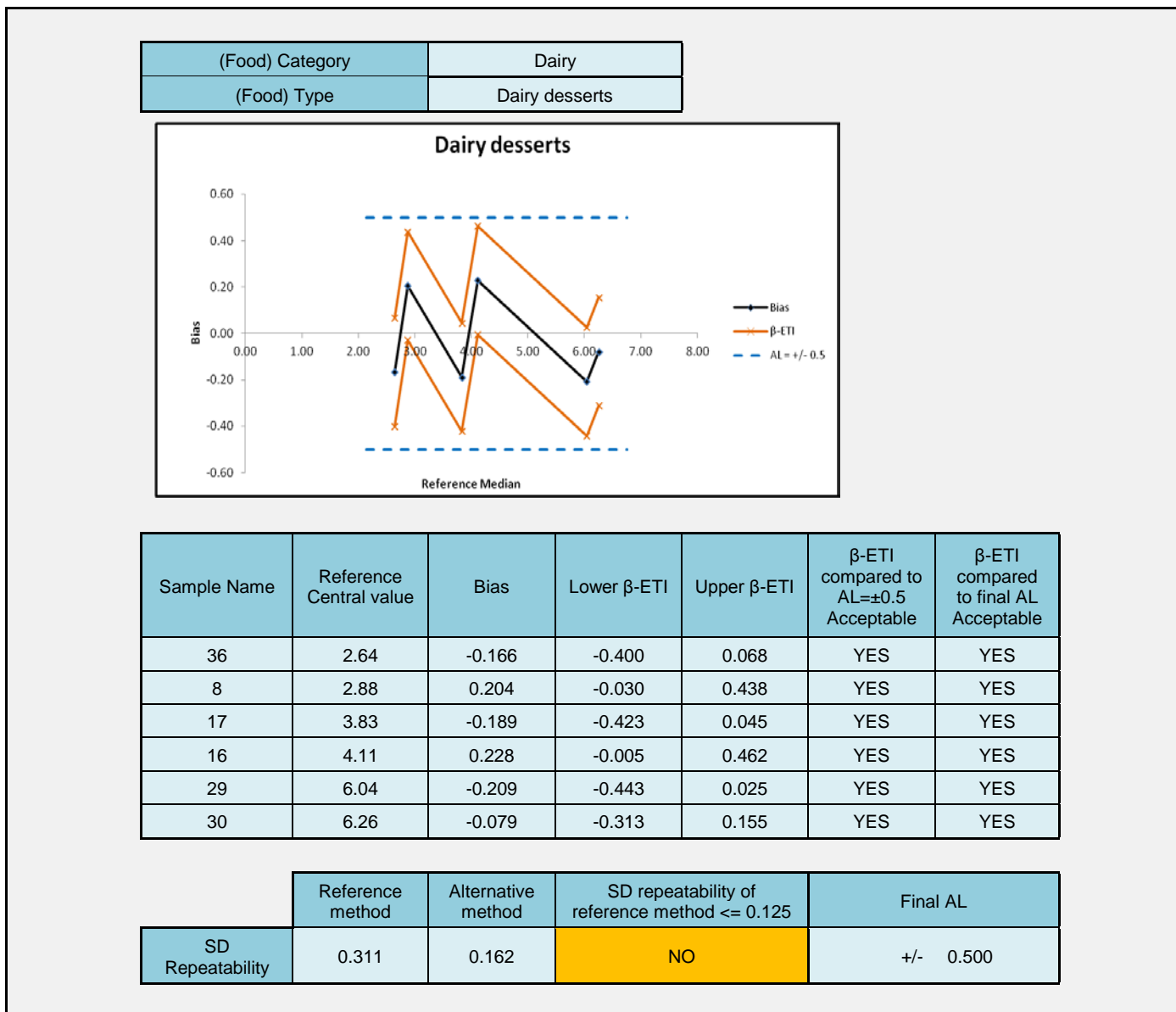


Figure 2b: Dried/rehydrated & low moisture products

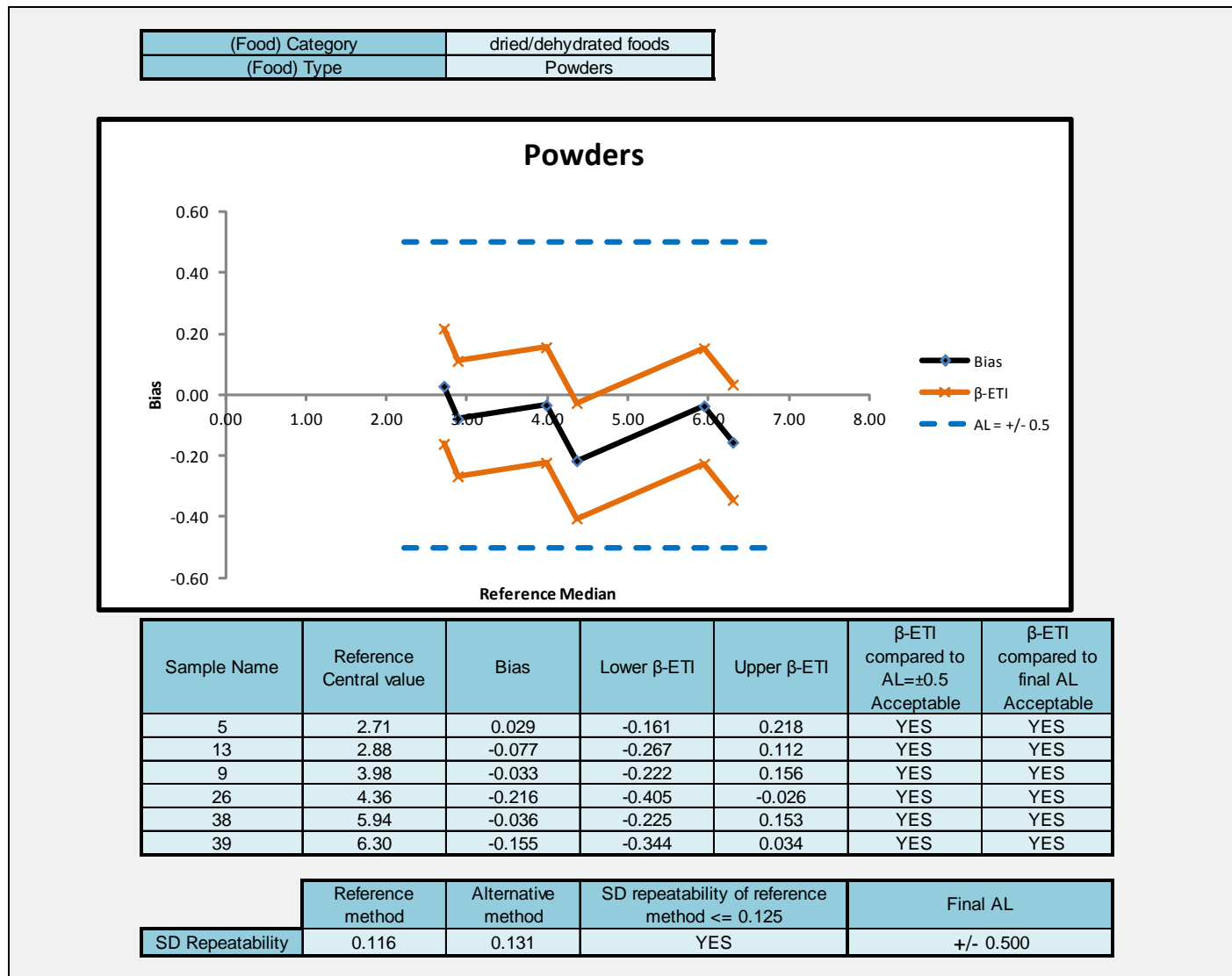


Figure 2c: Meat and poultry

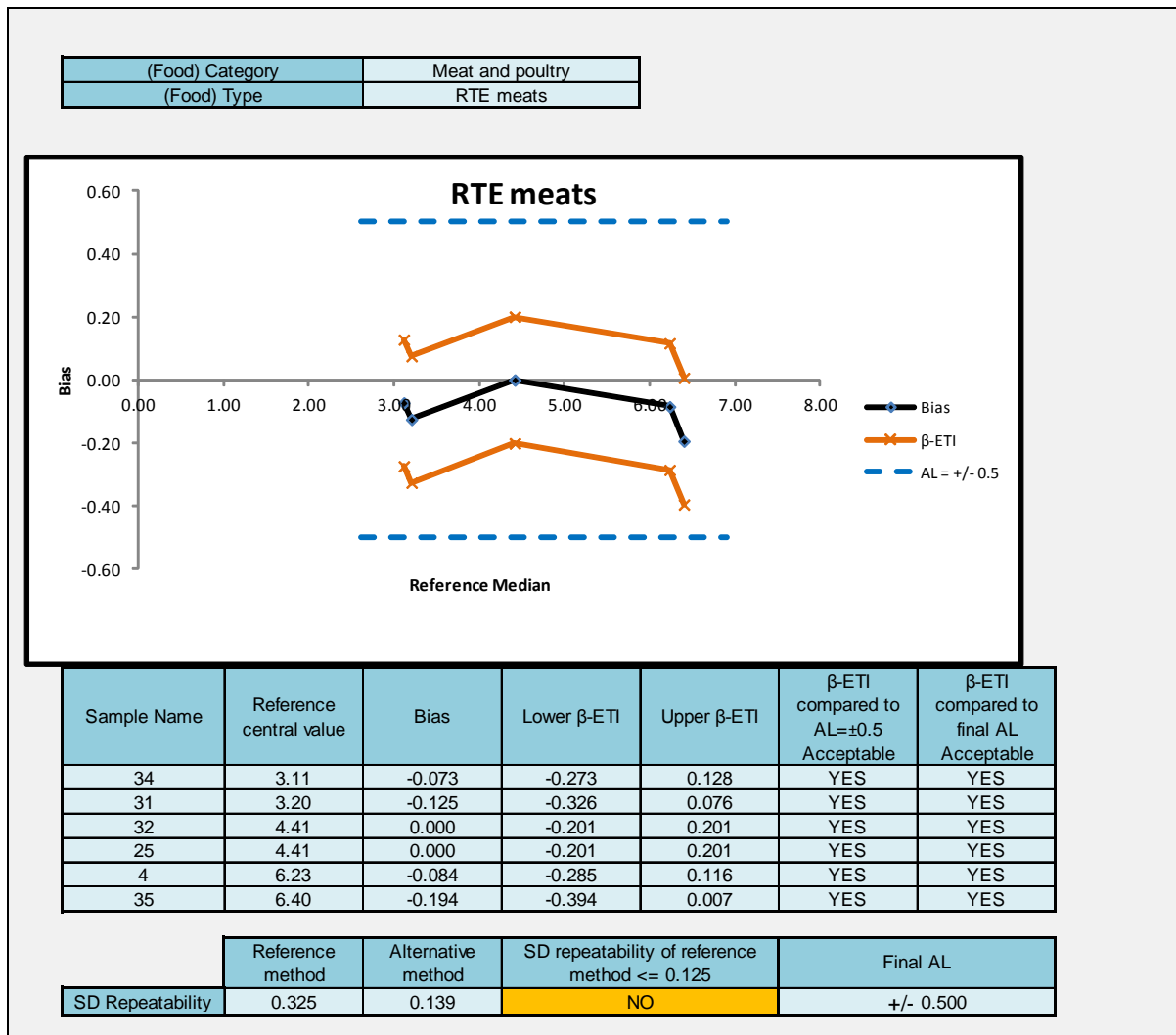


Figure 2d: Ready to eat foods

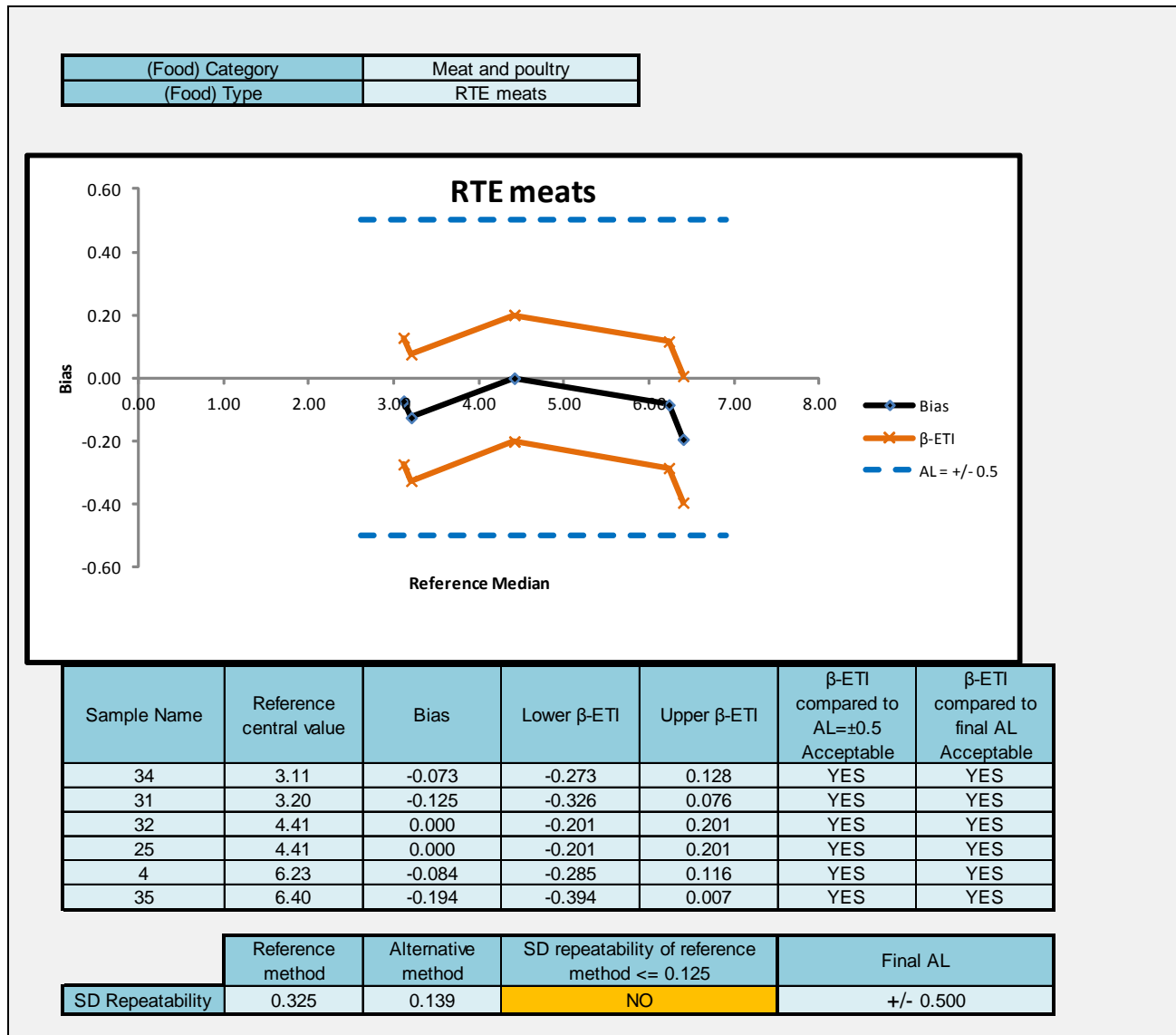
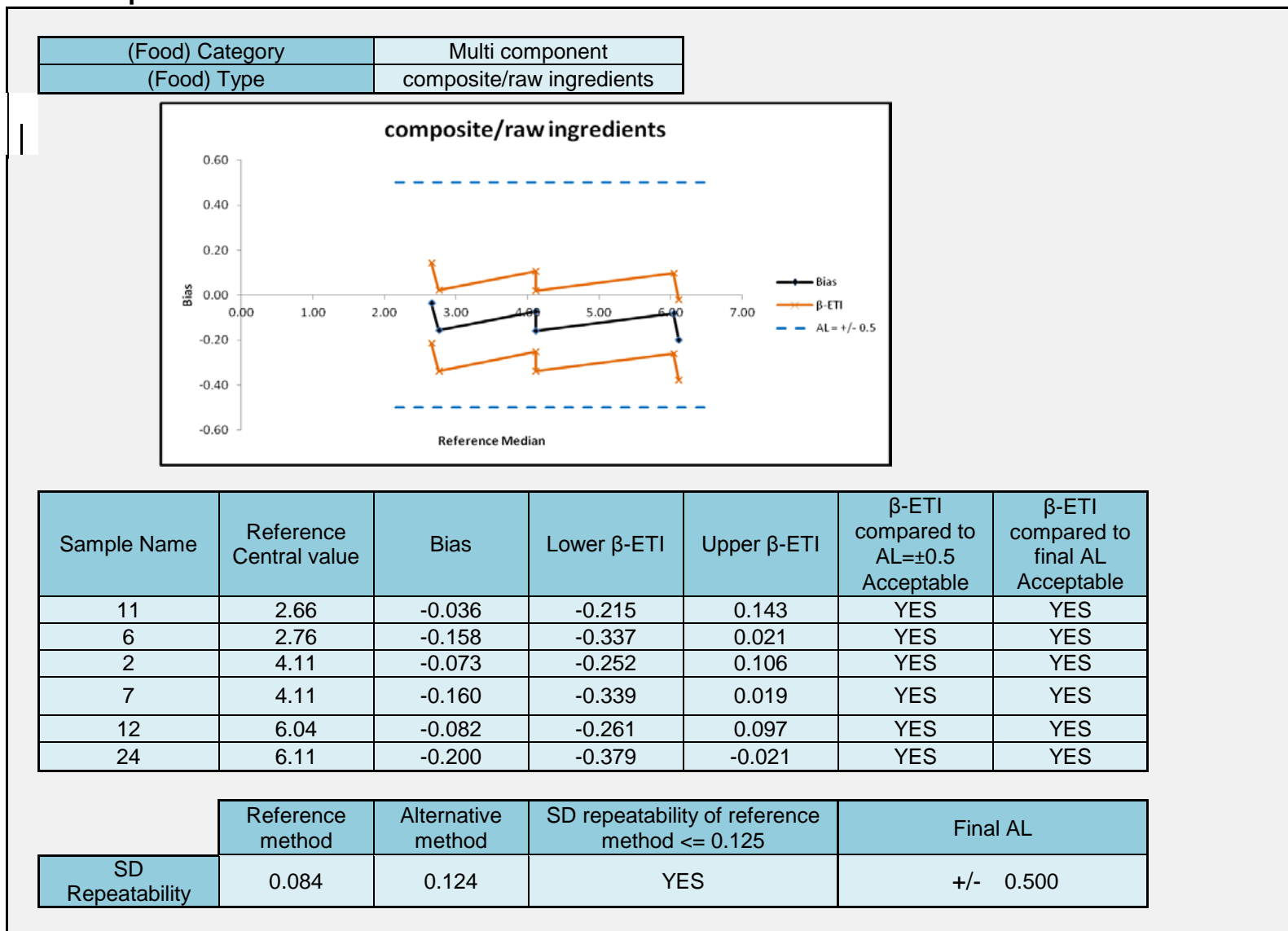


Figure 2e: Multi component foods



RESULTS OF THE COLLABORATIVE STUDY

Ten collaborative laboratories (as well as the Expert Laboratory) participated in the collaborative study. A single strain of *S.aureus* isolated from milk powder was incubated. Raw whole milk was used as matrix. Two milk samples remained uninoculated. For six samples, appropriated dilutions of the *S.aureus* culture were used to individually inoculated 2 x 20 ml milk samples at the lower (100 cfu/ml) middle (1000 cfu/ml) and higher (10000 cfu/ml) contamination levels. The results obtained are given in table 1.

Table 1. Statistical analysis of the ILS data according to the ISO spreadsheet

Accuracy profile				Reference method		
Study Name	XSA ILS analysis			Low	Medium	High
Date	Campden BRI					
Coordinator	22/12/2016					
Tolerance probability (beta)	80%	80%	80%			
Acceptability limit in log (lambda)	1.11	1.11	1.11			
Alternative method				Reference method		
Levels	Low	Medium	High	Low	Medium	High
Target value	3.100	4.061	5.102			
Number of participants (K)	10	10	10	10	10	10
Average for alternative method	2.677	3.798	4.897	3.100	4.061	5.102
Repeatability standard deviation (sr)	0.165	0.096	0.121	0.142	0.126	0.373
Between-labs standard deviation (sL)	0.277	0.145	0.336	0.178	0.196	0.309
Reproducibility standard deviation (sR)	0.323	0.174	0.358	0.228	0.233	0.485
Corrected number of dof	11.659	12.162	10.090	13.149	12.044	15.678
Coverage factor	1.417	1.412	1.435			
Interpolated Student t	1.359	1.355	1.371			
Tolerance interval standard deviation	0.3364	0.1810	0.3740			
Lower TI limit	2.220	3.552	4.384			
Upper TI limit	3.134	4.043	5.410			
Bias	-0.423	-0.263	-0.205			
Relative Lower TI limit (beta = 80%)	-0.880	-0.508	-0.718			
Relative Upper TI limit (beta = 80%)	0.034	-0.018	0.308			
Lower Acceptability Limit	-1.11	-1.11	-1.11			
Upper Acceptability Limit	1.11	1.11	1.11			
New acceptability limits may be based on reference method pooled variance						
Pooled repro standard dev of reference	0.337					

TRUE

Application of clause 6.2.3

Step 8: If any of the values for the β -ETI fall outside the acceptability limits, calculate the pooled average reproducibility standard deviation of the reference method.

Step 9: Calculate new acceptability limits as a function of this standard deviation.

TRUE
FALSE

Select ALL blue lines to draw the accuracy profile as illustrated in the worksheet "Graph Profile"



CONCLUSION:

According to the comparison and the collaborative study no statistical differences were found between the Compact Dry X-SA method and the reference method (ISO 6888-1:1999) for the enumeration of *Staphylococcus aureus*.

The observed profiles are within the 0.5 log AL or the recalculated AL limit calculated according to NordVal International Protocol /ISO16140-2:2016, and hence the alternative method is accepted as being equivalent to the reference method.